



Taq Polymerase

Thermostable DNA Polymerase
Thermus aquaticus, recombinant, *E. coli*

Cat. No.	Amount
TAQ_100KU	100 kunits
TAQ_1000KU	1.000 kunits

Unit Definition: One unit is defined as the amount of the enzyme required to catalyze the incorporation of 10 nmol of dNTP into an acid-insoluble form in 30 minutes at 70 °C using hering sperm DNA as substrate.

For *in vitro* use only!

Shipping: shipped on blue ice

Storage Conditions: store at -20 °C

Additional Storage Conditions: avoid freeze/thaw cycles

Shelf Life: 12 months

Form: liquid

Concentration: 5 units/μl

Description:

Taq Polymerase is recommended for routine PCR applications (up to 4 kb fragment length), high throughput PCR or genotyping. The buffer system guarantees robust and reliable amplification results in almost all PCR applications. The Crystal Buffer contains a well-balanced ratio of potassium-, ammonium- and magnesium-ions to ensure high specificity and minimal by-product formation without the need of additional optimization steps.

Ruby Buffer additionally contains gel loading buffer and an inherent red dye. The red dye allows an easy visual control during PCR set-up and in combination with the density reagent the direct loading of the reaction product into the gel.

The enzyme replicates DNA at 72 °C. It catalyzes the polymerization of nucleotides into duplex DNA in 5'→3' direction in the presence of magnesium. It also possesses a 5'→3' polymerization-dependent exonuclease replacement activity but lacks a 3'→5' exonuclease (proof-reading) activity.

Content:

Taq Polymerase (red cap)

5 units/μl Taq DNA Polymerase in 20 mM Tris-HCl, 100 mM KCl, 0.1 mM EDTA, 1 mM DTT, 0.5 % Tween-20, 0.5 % Nonidet P-40, 50 % (v/v) Glycerol, pH 8.0 (25°C)

Ruby Buffer (black cap)

10 x conc. complete PCR buffer containing 200 mM Tris-HCl, KCl, (NH₄)₂SO₄ and 20 mM MgCl₂, red tracking dye and density reagent for gel loading

Crystal Buffer (green cap)

10 x conc. complete PCR buffer containing 200 mM Tris-HCl, KCl, (NH₄)₂SO₄ and 20 mM MgCl₂



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Assay Set-Up:

Before starting, vortex all components thoroughly to ensure homogeneity.

Prepare a premix for the number of assays you need according to the following protocol:

comp.	cap	stock conc.	final conc.	1 assay @20 µl	1 assay @ 50 µl
PCR-grade Water	white			fill up to 10 µl	fill up to 30 µl
Ruby Buffer or Crystal Buffer	black or green	10x	1x	2 µl	5 µl
dNTP Mix / 10 mM #NU-1006	white	10 mM	200 µM	0.4 µl	1 µl
Taq Polymerase	red	5 units/µl	0.025 units/µl	0.1 µl	0.25 µl
primer mix or each primer		10 µM each primer	200 - 400 nM each primer	0.4-0.8 µl	1 - 2 µl
template /sample DNA				10 µl < 10 ng DNA	20 µl < 20 ng DNA

Select PCR tubes, stripes or plates as recommended for your cyclor model.

Aliquot premix into each well and add template DNA (or PCR-grade Water for negative controls).

Cycling Conditions:

Spin down the tubes/plate briefly to remove bubbles and place them into the cyclor.

initial denaturation	95 °C	2 min	1x
denaturation	95 °C	10 - 20 sec	25 - 35x
annealing ¹⁾	50 - 68 °C	10 - 20 sec	
elongation ²⁾	72 °C	20 sec - 4 min	

¹⁾The annealing temperature depends on the melting temperature of the primers used.

²⁾The elongation time depends on the length of the fragments to be amplified. A time of 1 min/kb is recommended.

Gel Loading and Down-Stream Applications:

Ruby Buffer includes a density reagent + tracking dye and allows the direct loading of the PCR products into a electrophoresis gel. For DNA detection / fluorescent DNA staining we recommend to use new generations dyes (i.g. SYBR DNA Stain) instead of the classical but highly mutagenic ethidium bromide.

Crystal Buffer is recommended for down-stream applications such as DNA sequencing, ligation, restriction digestion or where an analysis of the PCR product by absorbance or fluorescence excitation is required. For gel electrophoresis add gel loading buffer and fluorescent DNA stain before loading the PCR into the gel. Using pre-stained gels or post-run staining protocols is also possible.

Optimization of MgCl₂ concentration:

A final Mg²⁺ concentration of 2.0 mM is recommended in combination with Labeling Buffer. However, if an individual Mg²⁺ optimization is essential add 25 mM MgCl₂ stock solution as shown in the table below.

final MgCl ₂ conc.	20 µl final assay volume	50 µl final assay volume
2 mM	-	-
3 mM	0.8 µl	2.0 µl
4 mM	1.6 µl	4.0 µl
5 mM	2.4 µl	6.0 µl